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Research Article

Comparison of salivary Visfatin in patients with periodontitis and peri-implantitis in an Iranian population

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Abstract

Background. Visfatin (pre-B-cell colony-enhancing factor) is a salivary biomarker secreted from a variety of cells and is thought to have some proinflammatory and immune-modulating effects. This study compared salivary concentrations of visfatin in patients with chronic periodontitis and periimplantitis and healthy individuals.

Methods. In this cross-sectional and descriptive trial, patients were selected on the basis of inclusion and exclusion criteria. The whole saliva samples were collected; then according to the measured clinical parameters the patients were categorized to peri-, chronic periodontitis and periodontally healthy individuals. The concentrations of visfatin were evaluated using a standard ELISA kit. The salivary concentrations of visfatin were statistically analyzed using Kruskal-Wallis test. A probability value of less than 0.05 was considered significant.

Results. A total of 40 participants (21 females and 19 males) were enrolled in this study. The mean salivary visfatin concentrations in the periodontally healthy individuals, periimplantitis patients and chronic periodontitis patients were 23.97 ng/mL, 12.83 ng/mL and 11.95 ng/mL, respectively. However; visfatin levels were higher in healthy individuals compared to other groups No significant differences were found in salivary visfatin concentrations between the three groups.

Conclusion. Under the limitations of this study, no significant relationships were found regarding salivary concentrations of visfatin among periimplantitis and chronic periodontitis patients as compared to healthy individuals; however, more studies are required in this regard.

Key words: Visfatin, saliva, periimplantitis, chronic periodontitis.

Introduction

Chronic periodontitis and periimplantitis are inflammatory diseases resulting in irreversible loss of connective tissue and alveolar bone destruction. Leaving them untreated, tooth loss and implant failure may occur.

Periodontitis is one of the most prevalent diseases affecting more than 50% of the adult population and

its severe forms affect 11% of adults.

The increased demand for dental implants have resulted in an increase in the incidence of peri-implant diseases. The recent concern of individuals and practitioners is the 22% prevalence of periimplantitis, which is in need of greater attention.¹

Early detection of diseases with an emphasis on screening patients, as well as professional interventions, help prevent complications, improve treatment and control diseases.²

Identfying biomarkers related to inflamatory condictions may help determine the presence, risk and progression of periodontal diseases such as periodontitis and periimplantitis, and it may help transform biomarker technology from the labaratory to chariside activities.³ Inflammatory biomarkers can easily be assessed in saliva. Therefore, saliva samples provide a non-invasive diagnostic test to evaluate periodontal diseases.⁴ Some studies have reported the measurement of cytokines and biomarkers in inflammatory disorders, especially the relevant biomarkers of periodontitis, including adiponectin, resistin, leptin, visfatin, tumor necrosis factor alpha (TNF- α) and interleukin-6 (IL-6).⁵

Visfatin, originally named as pre-B-cell colonyenhancing factor and nicotinamide phosphoribosyl transferase, is a protein synthesized and secreted from adipose tissues and several cell types, including lymphocytes, bone marrow cells, hepatic cells, skeletal muscles, trophoblasts, and fetal membranes.⁶ This biomarker acts as a growth factor, cytokine, and enzyme and also induces destructive cytokines (IL-1b, TNF-a, and IL-6) in response to infection and inflammation.⁷

Few studies have evaluated gingival crevicular fluid (GCF) and serum levels of visfatin in periodontal diseases as in other inflammatory disorders. It has been demonstrated that visfatin levels positively correlated with the severity of periodontal disease.⁸⁻¹³ Salivary levels of visfatin also increase in patients suffering from periodontal inflammatory diseases as well.¹⁴⁻¹⁶

The aim of this study was to compare the salivary visfatin concentrations in patients with periimplantitis, periodontitis and healthy controls.

Methods

This case–control cross-sectional study was performed from March 2014 to March 2015. Patients visited with suspected periodontitis and patients who had received dental implants in the Department of Periodontics, Dental School, Shahid Beheshti University of Medical Sciences, were recalled and completed personal, medical and dental history questionnaires, and written consent was obtained. The study protocol was approved by the Ethics Committee of Shahid Beheshti University of Medical Sciences.

Inclusion criteria of the study were: age over 18 years, no known systemic disease and a minimum of 20 teeth

Exclusion criteria included a history of periodontal/peri-implant therapy during the past 2 years, a history of alcoholism and smoking; liver, kidney or salivary gland dysfunction; infectious diseases; inflammatory bowel disease; rheumatoid arthritis; granulomatous diseases; hypertension; diabetes; organ transplantation and cancer therapy. In addition, patients using glucocorticoids, cyclo-oxygenase inhibitors, bisphosphonates, antibiotics or immunosuppressant medications during the past 6 months, pregnant or lactating women, those needing antibiotics for infective endocarditis prophylaxis during dental procedures, those with acute illness symptoms (i.e. fever, sore throat, body aches or diarrhea), those wearing orthodontic appliances and those with an oral mucosal inflammatory condition (e.g. aphthous, lichen planus, leukoplakia or oral cancer) were excluded from the study.

Saliva Collection

Un-stimulated whole saliva (about 5 mL) samples were collected according to a modification of the Navazesh method between 10.00 AM and 12.00 PM.¹⁷

The patients were asked to swallow their saliva first, and then allow the saliva to drain passively for five minutes over the lower lip into a sterile tube. Collected salivary samples were immediately placed on ice and frozen at -40°C. The samples were analyzed within 6 months of collection.

Clinical Evaluation

Clinical measurements were performed at 6 points per tooth using a UNC probe and recorded in a chart by a periodontist. Diagnosis was made based on clinical parameters, including plaque index (PI), probing pocket depth (PPD), clinical attachment level (CAL) and bleeding index (BI).^{18,19}

The inclusion criteria consisted of patients with chronic periodontitis with at least five sites with CAL \geq 3 mm and PPD \geq 5 mm in a minimum of two teeth in at least two quadrants.²⁰

The periimplantitis group included patients with no history of periodontitis who had one or more implants with more than 12 months of loading. This condition was clinically confirmed by probing depth of >5 mm in at least one measurement with bleeding on probing and/or suppuration. There had to be radiographic observation of crestal bone loss in at least one site around the implant. At least 2 threads of implants had to be exposed due to crestal bone loss. Implant success index (ISI) ratings of VI, VI, I and VIII were included in this study (Table 1).²¹

(SL: soft tissue level, HL: hard tissue level, PPD: probing pocket depth, BOP: bleeding on probing, RBL: radiographic bone loss detected via long cone parallel peri-apical technique, +: tissue level located at or coronal to the reference line, -: level apical to the reference line).²¹

Patients with no history of periodontitis and clinical attachment loss, BI<0.1 and PPD mm in all sites were considered as the healthy group.²²

Biomarker Analysis

Each saliva sample (500 µL) was pipetted into a clean microcap tube and clarified by centrifugation at 10,000 g for 5 minutes. The supernatant was transferred to clean microcap tubes and used immediately enzyme-linked immunosorbent for an assav (ELISA). Concentrations of visfatin were determined using Human Visfatin (VISFATIN) ELISA Kit, according to the manufacturer's instructions. The results of visfatin assay were expressed as ng/mL for concentrations. All the laboratory tests were performed in the Biology Department of Shahid Beheshti University of Medical Sciences (Bioassay Technology Laboratory, Shanghai, China).

Statistical Analysis

Statistical analyses were performed using SPSS 18. Comparisons of visfatin levels between the groups were performed with Kruskal-Wallis test. P-value <

0.05 was considered statistically significant.

Results

A total of 40 participants (21 females and 19 males) were enrolled in this study. They consisted of 16 patients with chronic periodontitis (8 females and 8 males/mean age: 38.12 years), 12 patients with periimplantitis (7 females and 5 males/mean age: 47.91 years) and 12 healthy individuals (6 females and 6 males/mean age: 35.08 years). The mean salivary visfatin concentrations in the periodontally healthy individuals, periimplantitis patients and chronic periodontitis patients were 23.97, 12.83 and 11.95 ng/mL, respectively (Table 2). However, visfatin levels were higher in the healthy individuals compared to other groups (Figure 1).

No significant differences were found between salivary visfatin concentrations between the three groups (P=0.12).

Discussion

Various inflammatory biomarkers can be detected in both saliva and GCF.²³ While GCF sampling is somehow invasive, collection of saliva can be used for convenient diagnosis of oral diseases.²⁴

Visfatin, an inflammatory mediator which plays a role in increasing the secretion of TNF- α and IL-6 in monocytes, has been demonstrated to be increased by IL-1 β in chondrocytes; consequently, the result is an increase in paracrine/autocrine function, which would lead to higher levels of prostaglandin E2.²⁵

It has also been demonstrated that visfatin levels of serum increases in patients with chronic kidney disease. chronic obstructive pulmonary disease, Behcet's syndrome, appendicitis, metabolic syn-

Score	SL	HL	Clinical finding		
ISI I	SL+, PPD≤4mm, BOP-	HL+ Clinically healthy			
ISI II	SL+, PPD≤4mm, BOP+	HL+ Soft tissue information			
ISI III	SL+, PPD>4mm, BOP+	HL+	Deep soft tissue pocket		
ISI IV	SL+	HL-, RBL≤2mm(≤20%) Initiation of hard tissue breakdown			
ISI V	SL-	Hard tissue breakdown plus			
			Soft tissue recession		
ISI VI	SL+	HL-, RBL: 2-4mm(40%)	Notable hard tissue breakdown		
ISI VII	SL-	HL-, RBL: 2-4mm(40%) Notable hard tissue break			
			Plus soft tissue recession		
ISI VIII	-	RBL≥40% Severe bone loss			
ISI IX	-	Clinical mobility	Clinical failure		

Table 1. Implant success index (ISI))		
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Table 2. The diffusio	n index of salivary	y visfatin level i	n patients with	peri implantitis,	periodontitis and healthy
group					

Group	Number	Mean (ng/mL)	Med (ng/mL)	Min (ng/mL)	Max (ng/mL)	SEM (ng/mL)
Periimplantitis	12	12.83	12.78	0	26.45	3.21
Periodontitis	16	11.95	0	0	33.66	3.57
Healthy	12	23.97	19.83	8.77	43.61	2.99

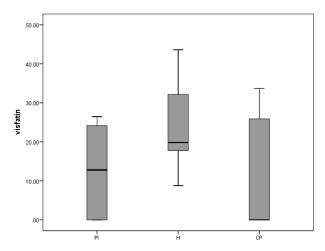


Figure 1. Salivary visfatin levels (ng/mL) in patients with periimplantitis (PI), periodontitis (P) and healthy individuals (H).

drome, inflammatory bowel disease and rheumatoid arthritis.^{7,26-31}

In periodontitis, visfatin is responsible for inflammatory reactions in periodontal structures and the consequent bone loss. It has been shown that microbial infection (presence of *Fusobacterium nucleatum*) has a controlling influence on production of visfatin.³²

This study was designed to evaluate salivary visfatin concentrations in 40 age- and sex-matched patients with periimplantitis and periodontitis and periodontally healthy individuals. The results showed no significant differences in salivary visfatin levels between these three groups. In addition, in some of the samples related to periimplantitis and periodontitis groups, no visfatin was detected.

(PI: Periimplantitis; P: Periodontitis; G: Gingivitis; H: Healthy)

As shown in Table 3, our findings are contrary to those in other studies; probable reasons and limitations are listed below:

1. The small sample size, which is due to two main reasons:

strict criteria for inclusion and exclusion; however,

this minimized the potential confounding factors and low incidence of periimplantitis

2. The presence of various standard classification systems approved for the diagnosis of disease.

Most studies consider a probing depth of >5 mm, bleeding on probing with or without discharging pus and evidence of bone loss as criteria to determine periimplantitis.³³ Probing depth alone is not an appropriate criterion and depends on factors such as the thickness of soft tissue, the depth of implant shoulders and the design of implant crest module. That's why we added it to the above criteria. Exposure of at least two threads of implant, which can be observed on radiograph, would provide a more accurate diagnosis of periimplantitis.

3. Different types of implants may be another reason for controversies in the results.

4. Different loading periods of implants

5. Difficulty of clinical diagnosis of initial gingivitis from healthy individuals. Therefore, a number of healthy people in the study might be in the early stages of gingivitis.

6. According to a study by Mamali et al, measurement of visfatin in saliva might not be reliable; therefore, further studies are necessary in order to confirm whole saliva as a proper fluid for the evaluation of visfatin.34

7. In this study, the time of collecting saliva samples differed from other studies. Not to interfere in induction of biomarkers, the sampling was conducted before measuring PPD and BOP.

Further studies are needed to evaluate the effects of the factors we excluded from the study. In order to consider salivary visfatin as an inflammatory biomarker of periodontitis, further longitudinal studies are necessary, using a larger sample size in different populations.

Conclusion

Under the limitations of this study, no significant relationships were found in salivary concentrations of visfatin between periimplantitis and chronic peri-

Table 3. Studies on the level of visfatin in periodontal diseases

Studies on visfatin	Number of patients	Groups	Sample	Analysis by	Result
Paradeep et al 2011	40	CP,G,H	GCF, Serum	ELISA	increased in GCF and serum with the severity of disease from healthy to gingivitis to periodontitis groups
Raghavendra et al 2012	30	CP, Treated CP, H	GCF, Serum	ELISA	decreased after non-surgical periodontal treatment
Paradeep et al 2012	30	DM+CP,CP ,H	GCF, Serum	ELISA	increased in both serum and GCF in individuals with t2 DM with CP
Alizadeh Tabari et al 2013	40	CP, H	Saliva	ELISA	significantly higher in periodontitis group
Ozcan et al 2014	72	СР, G, H	Saliva	ELISA	higher in patients with gingivitis and periodontitis compared to healthy subjects, gingivitis = periodontitis
Alizadeh Tabari et al 2015	40	CP, Treated Saliva, CP, H	Saliva	ELISA	reduced after non-surgical periodontal therapy to the levels comparable with those found in healthy individuals.
Present study	40	PI, CP, H	Saliva	ELISA	higher in healthy individuals; no significant differences were found in between the three groups

odontitis patients and healthy controls.

Competing interests

There is no conflict of interests in this work.

References

- 1. Tonetti MS, Chapple IL, Jepsen S, Sanz M. Primary and secondary prevention of periodontal and peri-implant diseases: Introduction to, and objectives of the 11th European Workshop on Periodontology consensus conference. Journal of clinical periodontology. 2015;42 Suppl 16:S1-4. doi: 10.1111/jcpe.12382
- Lee YH, Wong DT. Saliva: an emerging biofluid for early detection of diseases. American journal of dentistry. 2009;22(4):241-8. doi: 10.1111/j.1601-0825.2010.01773.x
- Syndergaard B, Al-Sabbagh M, Kryscio RJ, Xi J, Ding X, Ebersole JL, et al. Salivary biomarkers associated with gingivitis and response to therapy. Journal of periodontology. 2014;85(8):e295-303. doi: 10.1902/jop.2014.130696
- Miller CS, Foley JD, Bailey AL, Campell CL, Humphries RL, Christodoulides N, et al. Current developments in salivary diagnostics. Biomarkers in medicine. 2010;4(1):171-89. doi: 10.2217/bmm.09.68
- Fantuzzi G. Adipose tissue, adipokines, and inflammation. The Journal of allergy and clinical immunology. 2005;115(5):911-9; quiz 20. doi: 10.1016/j.jaci.2005.02.023
- Fukuhara A, Matsuda M, Nishizawa M, Segawa K, Tanaka M, Kishimoto K, et al. Visfatin: a protein secreted by visceral fat that mimics the effects of insulin. Science (New York, NY). 2005;307(5708):426-30. doi: 10.1126/science.1097243
- Moschen AR, Kaser A, Enrich B, Mosheimer B, Theurl M, Niederegger H, et al. Visfatin, an adipocytokine with proinflammatory and immunomodulating properties. Journal of immunology (Baltimore, Md : 1950). 2007;178(3):1748-58. doi: 10.4049/jimmunol.178.3.1748
- Jia SH, Li Y, Parodo J, Kapus A, Fan L, Rotstein OD, et al. Pre-B cell colony-enhancing factor inhibits neutrophil apoptosis in experimental inflammation and clinical sepsis. The Journal of clinical investigation. 2004;113(9):1318-27. doi: 10.1172/jci200419930
- Otero M, Lago R, Gomez R, Lago F, Dieguez C, Gomez-Reino JJ, et al. Changes in plasma levels of fat-derived hormones adiponectin, leptin, resistin and visfatin in patients with rheumatoid arthritis. Annals of the rheumatic diseases. 2006;65(9):1198-201. doi: 10.1136/ard.2005.046540
- Koczan D, Guthke R, Thiesen HJ, Ibrahim SM, Kundt G, Krentz H, et al. Gene expression profiling of peripheral blood mononuclear leukocytes from psoriasis patients identifies new immune regulatory molecules. European journal of dermatology : EJD. 2005;15(4):251-7. doi: 10.1016/j.molimm.2013.05.127
- 11. Pradeep AR, Raghavendra NM, Prasad MV, Kathariya R, Patel SP, Sharma A. Gingival crevicular fluid and serum visfatin concentration: their relationship in periodontal health and disease. Journal of periodontology. 2011;82(9):1314-9. doi: 10.1902/jop.2011.100690
- 12. Pradeep AR, Raghavendra NM, Sharma A, Patel SP, Raju A, Kathariya R, et al. Association of serum and crevicular visfatin levels in periodontal health and disease with type 2

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diabetes mellitus. Journal of periodontology. 2012;83(5):629-34. doi: 10.1902/jop.2011.110272

- 13. Raghavendra NM, Pradeep AR, Kathariya R, Sharma A, Rao NS, Naik SB. Effect of non surgical periodontal therapy on gingival crevicular fluid and serum visfatin concentration in periodontal health and disease. Disease markers. 2012;32(6):383-8. doi: 10.1155/2012/828049
- 14. Ghallab NA, Amr EM, Shaker OG. Expression of Leptin and Visfatin in Gingival Tissues of Chronic Periodontitis With and Without Type 2 Diabetes Mellitus: A Study Using Enzyme-Linked Immunosorbent Assay and Real-Time Polymerase Chain Reaction. Journal of periodontology. 2015;86(7):882-9. doi: 10.1902/jop.2015.140434
- 15. Abolfazli N, Jabali S, Saleh Saber F, Babaloo Z, Shirmohammadi A. Effect of Non-surgical Periodontal Therapy on Serum and Salivary Concentrations of Visfatin in Patients with Chronic Periodontitis. Journal of dental research, dental clinics, dental prospects. 2015;9(1):11-7. doi: 10.15171/joddd.2015.003
- 16. Tabari ZA, Azadmehr A, Nohekhan A, Naddafpour N, Ghaedi FB. Salivary visfatin concentrations in patients with chronic periodontitis. Journal of periodontology. 2014;85(8):1081-5. doi: 10.1902/jop.2013.130388
- Navazesh M. Methods for collecting saliva. Annals of the New York Academy of Sciences. 1993;694:72-7. doi: 10.1111/j.1749-6632.1993.tb18343.x
- 18. Silness J, Loe H. PERIODONTAL DISEASE IN PREGNANCY. II. CORRELATION BETWEEN ORAL HYGIENE AND PERIODONTAL CONDTION. Acta odontologica Scandinavica. 1964;22:121-35. doi: 10.3109/00016356408993968
- 19. Ainamo J, Bay I. Problems and proposals for recording gingivitis and plaque. International dental journal. 1975;25(4):229-35. doi: 10.1111/j.1600-0722.1975.tb00414.x
- 20. Al-Sabbagh M, Alladah A, Lin Y, Kryscio RJ, Thomas MV, Ebersole JL, et al. Bone remodeling-associated salivary biomarker MIP-1alpha distinguishes periodontal disease from health. Journal of periodontal research. 2012;47(3):389-95. doi: 10.1111/j.1600-0765.2011.01445.x
- Kadkhodazadeh M, Esfahrood ZR, Amid R, Zarnegarnia P. Comparison of the acceptability of a new scoring system with Misch's classification for dental implant success determination. Journal of long-term effects of medical implants. 2012;22(1):85-93. doi: 10.1615/jlongtermeffmedimplants.v22.i1.90
- 22. Armitage GC. Development of a classification system for periodontal diseases and conditions. Annals of periodontology / the American Academy of Periodontology. 1999;4(1):1-6. doi: 10.1902/annals.1999.4.1.1
- 23. Giannobile WV, Beikler T, Kinney JS, Ramseier CA, Morelli T, Wong DT. Saliva as a diagnostic tool for periodontal disease: current state and future directions. Periodontology 2000. 2009;50:52-64. doi: 10.1111/j.1600-0757.2008.00288.x
- Buduneli N, Kinane DF. Host-derived diagnostic markers related to soft tissue destruction and bone degradation in periodontitis. Journal of clinical periodontology. 2011;38 Suppl 11:85-105. doi: 10.1111/j.1600-051x.2010.01670.x
- 25. Bae YH, Bae MK, Kim SR, Lee JH, Wee HJ, Bae SK. Upregulation of fibroblast growth factor-2 by visfatin that promotes endothelial angiogenesis. Biochemical and biophysical research communications. 2009;379(2):206-11. doi: 10.1016/j.bbrc.2008.12.042

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- 26. Axelsson J, Witasp A, Carrero JJ, Qureshi AR, Suliman ME, Heimburger O, et al. Circulating levels of visfatin/pre-B-cell colony-enhancing factor 1 in relation to genotype, GFR, body composition, and survival in patients with CKD. American journal of kidney diseases : the official journal of the National Kidney Foundation. 2007;49(2):237-44. doi: 10.1053/j.ajkd.2006.11.021
- 27. Liu X, Ji Y, Chen J, Li S, Luo F. Circulating visfatin in chronic obstructive pulmonary disease. Nutrition (Burbank, Los Angeles County, Calif). 2009;25(4):373-8. doi: 10.1016/j.nut.2008.09.008
- Sezen H, Okumus S, Pehlivan Y, Dilli I, Tarakcioglu M, Onat AM. Visfatin levels in Behcet's disease. Inflammation. 2012;35(2):405-8. doi: 10.1007/s10753-011-9328-2
- Isman FK, Zulfikaroglu B, Acar A, Koc M, Tez M, Kucur M. Serum visfatin levels in acute appendicitis. International journal of surgery (London, England). 2010;8(3):259-60. doi: 10.1016/j.ijsu.2010.02.006
- 30. Filippatos TD, Derdemezis CS, Kiortsis DN, Tselepis AD, Elisaf MS. Increased plasma levels of visfatin/pre-B cell colony-enhancing factor in obese and overweight patients

with metabolic syndrome. Journal of endocrinological investigation. 2007;30(4):323-6. doi: 10.1007/bf03346300

- Brentano F, Schorr O, Ospelt C, Stanczyk J, Gay RE, Gay S, et al. Pre-B cell colony-enhancing factor/visfatin, a new marker of inflammation in rheumatoid arthritis with proinflammatory and matrix-degrading activities. Arthritis and rheumatism. 2007;56(9):2829-39. doi: 10.1002/art.22833
- 32. Nogueira AV, Nokhbehsaim M, Eick S, Bourauel C, Jager A, Jepsen S, et al. Regulation of visfatin by microbial and biomechanical signals in PDL cells. Clinical oral investigations. 2014;18(1):171-8. doi: 10.1007/s00784-013-0935-1
- Heitz-Mayfield LJ. Peri-implant diseases: diagnosis and risk indicators. Journal of clinical periodontology. 2008;35(8 Suppl):292-304. doi: 10.1111/j.1600-051x.2008.01275.x
- 34. Mamali I, Roupas ND, Armeni AK, Theodoropoulou A, Markou KB, Georgopoulos NA. Measurement of salivary resistin, visfatin and adiponectin levels. Peptides. 2012;33(1):120-4. doi: 10.1016/j.peptides.2011.11.007