Journal of Advanced Periodontology & Implant Dentistry

Research Article

Effect of IL-36γ expression on chronic periodontitis with an increase in NF-KB signaling pathway

Amir Reza Babaloo¹ • Adileh Shirmohammadi¹ • Shima Ghasemi² • Siamak Sandoghchian³ • Sahel

Parvan⁴* • Nazanin Fathi⁵

¹Department of Periodontics, Faculty of Dentistry, Tabriz University of Medical Sciences, Tabriz, Iran ²Department of Prosthodontics, Faculty of Dentistry, Tabriz University of Medical Sciences, Tabriz, Iran ³Department of Immunology, Faculty of Medicine, Tabriz University of Medical Sciences, Tabriz, Iran ⁴Dentist in Private Practice, Tabriz, Iran ⁵Faculty of Dentistry, Tabriz University of Medical Sciences, Tabriz, Iran *Corresponding Author; E-mail: parvan_s1372@yahoo.com

> Received: 12 May 2019; Accepted: 13 August 2019 J Adv Periodontal Implant Dent 2019;11(1):28–33| doi:10.15171/japid.2019.005 This article is available from https://japid.tbzmed.ac.ir/

© 2019 The Authors. This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Abstract

Background. An inappropriate inflammatory response is the cause of many common diseases, especially periodontitis. Considering that no studies have been carried out to investigate the effect of IL-36γ on chronic periodontitis, this study aimed to investigate the inflammatory mechanism of IL-36γ by stimulating macrophage cells using NF-KB pathway.

Methods. This experimental study was performed on 50 healthy individuals and 50 subjects with chronic periodontitis. In this study, macrophage cells were extracted first, and then RNA was isolated from all the samples using TRIzol method. Subsequently, the rate of IL-36 γ gene expression was analyzed and compared using real-time PCR technique. Additionally, immunofluorescence (IF) technique was used to investigate the rate of inflammation. The rate of NF-Kb expression was also measured via western blot technique. Finally, statistical analysis of the samples was carried out using appropriate statistical methods with SPSS 17.

Results. The results showed that the rate of IL-36 γ expression in subjects with periodontitis was higher compared to healthy subjects (P<0.05). Moreover, the results showed that following treatment of cells with TLR4, the rate of IL36 γ expression increased significantly, especially during the 12-hour period after treatment.

Conclusion. This indicates that after stimulating the TLR pathways, the rate of IL-36 γ expression will probably increase.

Key words: Chronic periodontitis, Inflammation, TLR4, IL-36y, NF-KB.

Introduction

Periodontal pathogens are necessary for the onset of periodontal diseases. However, the prevalence of the disease and the severity of tissue destruction highly depend on the nature of host-microbe interactions. Host response to periodontal infections needs the expression of some bioactive factors such as proinflammatory and anti-inflammatory cytokines, growth factors and enzymes due to the activation of multiple signaling pathways. In fact, most of the Inflammation includes some biochemical and cellular changes. Inappropriate inflammatory response is the cause of many common diseases such as periodontitis. Inflammatory cells are activated by cytokines, bioactive molecules, and MAMPs in the area and regulate the activity of other cells by producing other inflammatory mediators and affecting the hemostasis of mineral and non-mineral tissues in the periodontium. The cytokines responsible for the initial response to microbial attack include IL-6, IL-1 β , IL-1 α and TNF- α . The members of interleukin-1 family as the effective molecules in the innate immune system are the vital regulators of various inflammatory disorders and all immune cells express or are under the influence of the members of IL-1 family.^{2,3}

IL-36y regulates IL-1ß levels through NF-Kb signaling pathway and the factors are regulated by MAPK such as c-jun by binding to IL-36R receptor. In addition, IL-36y activates MAPK and NF-kB signaling pathways in epithelial cells.^{4,5} TLRs exist on the cells of innate immune system (cells involved in the first line of defense), including neutrophils, monocytes/macrophages, and epithelial cells. Such cells, playing a significant role in detecting and killing microorganisms, indicate different TLRs which cause different immune responses to a specific pathogen.^{6,7} Studies indicated that IL-36y works as a proinflammatory cytokine in inflammation.⁸ Such cytokines are able to stimulate the performance of Th17 by inducing the relevant cytokines.⁹ A study showed that macrophages and dendritic cells derived from human monocytes that expression of Th-17 chemokine could create inflammatory responses via IL-36y. In addition, the lipopolysaccharide (LPS) derived from E. coli and P. gingivalis significantly expresses IL-36y, which is different for IL-36a and IL-36B. As it was mentioned, these factors affect the role of IL-36y in inflammations while its role in the inflammation of chronic periodontitis has not been studied yet.^{10,11}

Studies on IL-36 γ indicated that TLR4 can increase IL-36 γ expression and can activate the signaling pathway of nuclear factor – KB (NF-KB) and MAPK (mitogen activated protein kinase).¹² Since no study has been conducted on the effect of IL-36 γ with the assessment of the effect of NF-KB factor on chronic condition of periodontitis, the present study investigated the effect of IL-36 γ expression on chronic periodontitis through an increase in NF-KB signaling pathway.

Methods

In this study, all the laboratory stages were conducted in the Immunology Laboratory of Immunology Research Center, Tabriz University of Medical Sciences. First the gingival tissues derived from a sub-marginal flap were prepared and then sent to the Immunology Laboratory in normal saline solution. Then, the macrophages were isolated from the tissue samples using the MACS (magnetic activated cell sorting) technique. In this way, the cells were isolated by specific nanoparticles which contain specific antibodies of CD14 and CD16. After the isolation, the cells were cultured in 1640 RPMI culture medium with 1% erythromycin.

Evaluation of Gene Expression

First, the cells were stimulated by TLR4 proteins at 2-, 4-, 8-, 12- and 24-hour intervals. Then, the cell mRNA was extracted by TRIzol technique and IL- 36γ level in macrophages was evaluated in a real-time PCR device (Rotogen Company). In this method, β -actin gene was selected as control. In addition, some part of the tissue was measured directly with IL- 36γ and β -actin primers through real-time PCR technique.

Immunofluorescence Technique

In order to prove inflammation, the samples were placed in a blocking buffer for one hour after paraffinization of the cells. The primary antibodies anti-CD4 and anti-IL-17 were placed at room temperature for two hours. After the rinse, the secondary antibody was added for one hour. Then, 50 μ L of hochest was poured onto the slides after incubation and studied under an immunofluorescence microscope.

Western Blot Technique

The tissues were electrophoresed on 12% SDS Page gel and then transferred to PVDF transfer plates. The plates were blocked with 5% skimmed milk with 1% Tween in PBS for one hour at room temperature and then incubated for one night by anti-TLR4 and anti-NF-KB purchased from Abcam Company. The effect was measured by Amercontrol software.

The Design of Primers by a Computer

In order to facilitate the primer design process and minimize the required time, a computer software program such as Oligo 7 can be used to design and blast the primers. Many types of computer software programs are used to search the sequence of an appropriate primer region and select the best primer pairs with favorable features.

30 Babaloo et al.

Real-time PCR

All real-time PCR reactions were performed on (Rotor Gene TM 6000 (Corbett) device. The relevant reactions were performed in three steps. The standard curve was drawn based on the logarithm of the horizontal axis of cDNA concentration and vertical axis of threshold cycles for each gene. The PCR yield was determined for each gene based on the standard curve. From the non-cDNA sample, 'no template control' (NTC) was used as a negative control. An amplification curve was drawn for each reaction. The analysis was conducted according to the comparison of the threshold cycles of patients having chronic periodontitis to healthy samples.

The research results were reported using descriptive statistics (mean \pm standard deviation and percentage) to compare the level of IL-36 γ expression. ANOVA was used to compare this amount in two pathways at different times. Statistical analysis was conducted using SPSS 17 and the significance level was defined at P<0.05.

Results

This study was experimental and 100 subjects were included in the study, of which 50 subjects were in the healthy group, with 50 in the patient group.

As can be observed in Figure 1, the percentage of expression was 17.3% in the healthy group, with 33.6% in the patient group. The results indicated that that expression of IL-36 γ in subjects with periodontitis was

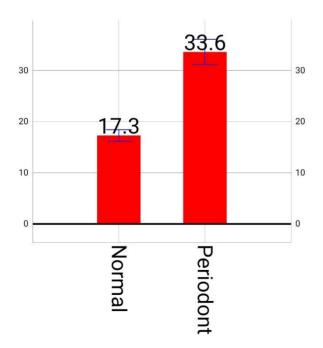


Figure 1. Comparison of gene expression between the two groups.

significantly different from the healthy subjects (P<0.001). This significant difference was due to the presence of inflammation in the subjects with periodontitis (Figure 1).

Comparison of gene expression in TLR4-treated cells was investigated at 2, 4, 8, 12 and 24 hours. The results showed that IL-36 γ gene expression increased with an increase in the duration of treatment with TLR4 and this increase peaked 12 hours later (48.3%). This amount decreased to 30.2% after 24 hours due to its reduced effectiveness. Such a difference in gene expression was reported significant at different times according to the results of a repeated measurement test (P<0.001) (Figure 2).

In this study, the amount of Nf-KB gene expression in patients with periodontitis was evaluated in IL-36γcarrying cells. As shown in Figure 3, Nf-KB in the periodontitis group increased in comparison to the sham group. In other words, IL-36γ can increase transcription by enhancing TLR4 pathway through MYD88 proteins as well as increasing the JAK and STAT pathways. Nf-KB levels increased with an increase in inflammatory cytokines (Figure 3).

Discussion

Chronic periodontitis is one of the infectious diseases caused by inflammation in the supporting tissues of teeth, leading to the loss of attachments and the alveolar bone.¹³ Many studies have considered the role of inflammatory and immune responses, especially

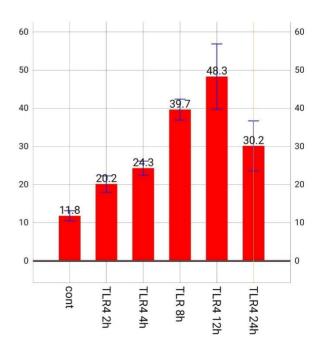


Figure 2. Comparison of TLR4-treated gene expression at different time intervals.

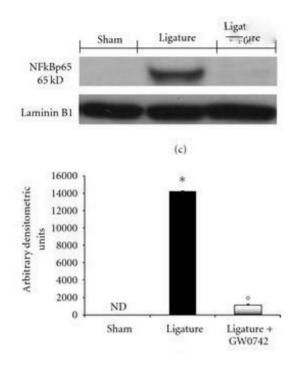


Figure 3. Comparison of Nf-KB gene expression to the sham group.

humoral immune responses, in the pathogenesis of this disease in recent years. These studies indicated that factors such as the complement system, collagenase and inflammatory cytokines, including the gene family of IL-1, TNF- α and IL-6, as important factors involved in the inflammatory system, are of great importance.¹⁴⁻¹⁶

In particular, IL-36 γ is one of the strongest inflammatory cytokines playing a main role in the reaction of host to gingivitis. Interestingly, the expression of IL-36 γ affects other cytokines of the IL-36 family. The cause of this effect is still unknown as how the effects of all the three various types of IL-36 occur through an IL-36R receptor. However, it might have different kinds of bioavailability in vivo, which might result in different inflammatory responses with different severities and durations.⁵

The results of this study revealed that IL-36 γ expression in the periodontitis group was significantly higher in comparison to the healthy group. In a study by Kurşunlu et al¹⁷ on the levels of IL-36 γ expression in periodontal patients, the results indicated no significant differences between different groups of invasive and chronic periodontitis and healthy subjects. However, IL-36 γ values in the chronic periodontitis group were higher than other groups, which is different from the results of this study. However, the values for IL-36 β in invasive periodontitis samples were more than the other samples and there was a statistically

Effect of IL-36y Expression on Chronic Periodontitis 31

significant difference. The results of this study demonstrated that cytokines such as IL-36y, in contrast to the family of IL-1 cytokines (the role of these proteins was mentioned in the progression of periodontal disease in previous studies) have no role in the pathogenesis of periodontal disease.¹⁸ Such results indicated that IL-36y cannot activate signaling functions related to IL-1 and likely act through alternative signaling pathways.¹⁹ In addition, the results of other studies showed that the production of these new molecules by inflammatory cells is low.^{5,20} The results of a study by Huynh et al^{21} indicated that IL-36 γ applies inflammatory responses through dendritic cells derived from monocytes as well as macrophages that includes the expression of neutrophil chemokines such as IL-8, CXCVL1, IL-17 and CCL20, IL-36v is similarly stimulated in the oral epithelial cells. IL-17 not only stimulates the expression of significant regulators which attract neutrophils but also leads to IL-36y expression in epithelial cells.

In addition, the results related to gene expression in TLR4-treated cells in the present study indicated that IL-36 γ gene expression increased with an increase in the duration of treatment by TLR4 and such an increase peaked 12 hours later. However, this amount decreased after 24 hours. The results of a study by Sara et al²² revealed that TLRs expression in chronic periodontitis significantly increased compared to healthy and gingivitis patients and TLR4 expression increased more than TLR2.22 Another study by Yamaguchi et al²³ investigated the effect of gingival plaque on the production of inflammatory cytokines by inducing TLR2 and TLR4 in peripheral blood mononuclear cells. The results indicated that levels of inflammatory cytokines, including TNFa, IL-6 and IL-8, were related to the ability of plaque to induce TLR4 pathway. Furthermore, it was found that cytokine production was inhibited by using TLR4 antagonists. It has been recently found that IRF6 (IFN regulatory

factor) is the key regulator for the expression of IL-36 γ through oral epithelial cells in response to gingivitis. IRF6 plays a critical role in regulating inflammatory cytokines caused by TLR2. Oral epithelium cells as the first cells encountering gingivitis play a significant role in oral periodontium.²⁴ The results of some studies indicated that IRF6 stimulates the expression of IL-1 family of inflammatory cytokines like IL-36 γ when oral epithelial cells encounter gingivitis.²¹

Conclusion

Based on the results, expression of IL-36 γ increased in subjects with periodontitis compared to healthy

32 Babaloo et al.

subjects. In addition, it was found that the level of IL-36 γ expression significantly increased after treating the cells with TLR4. The maximum amount of it was related to 12 hours after the treatment, indicating that the expression of IL36 γ increased by stimulating TLR pathways.

Authors' Contributions

ARB and ASh carried out surgeries and prepared the samples. SS was responsible for laboratory procedures. SP and SG were responsible for case selection, screening, coordination and preparation of the proposal; SP and NF prepared the manuscript.

Acknowledgments

None.

Funding

This study was financially supported by the Research Council of Tabriz University of Medical Sciences.

Competing Interests

The authors declare no competing interests with regards to the authorship and/or publication of this article.

Ethics Approval

This study was approved by the Ethics Committee of Tabriz University of Medical Sciences under the code IR.TBZMED.REC.1397.152

References

- 1. Flemmig TF. Periodontitis. Ann Periodontol. 1999;4(1):32-7. doi:10.1902/annals.1999.4.1.32
- 2. Dinarello CA. Interleukin-1 in the pathogenesis and treatment of inflammatory diseases. Blood .2011;117(14):3720-32. doi: 10.1182/blood-2010-07-273417
- Dinarello CA. Immunological and inflammatory functions of the interleukin-1 family. Annu. Rev. Immunol. 2009;27:519-50. doi: 10.1146/annurev.immunol.021908.132612
- Debets R, Timans JC, Homey B, Zurawski S, Sana TR, Lo S, et al. Two novel il-1 family members, il-1δ and il-1ε, function as an antagonist and agonist of nf-κb activation through the orphan il-1 receptor-related protein 2. J Immunol. 2001;167(3):1440-6.
- Towne JE, Garka KE, Renshaw BR, Virca GD, Sims JE. Il-1f6, il-1f8 and il-1f9 signal through il-1rrp2 and il-1racp to activate the pathway leading to nfkb and mitogen-activated protein kinases. J Biol Chem. 2004;279(14):13677-88. doi: 10.1074/jbc.M400117200
- Hayashi F, Means TK, Luster AD. Toll-like receptors stimulate human neutrophil function. Blood 2003;102(7):2660-9. doi:10.1182/blood-2003-04-1078
- Kusumoto Y, Hirano H, Saitoh K, Yamada S, Takedachi M, Nozaki T, et al. Human gingival epithelial cells produce chemotactic factors interleukin-8 and monocyte chemoattractant protein-1 after stimulation with porphyromonas gingivalis via toll-like receptor 2. *J* Periodontol . 2004;75(3):370-9. doi:10.1902/jop.2004.75.3.370

- Ramadas RA, Ewart SL, Medoff BD, LeVine AM. Interleukin-1 family member 9 stimulates chemokine production and neutrophil influx in mouse lungs. Am J Respir *Cell* Mol Biol. 2011;44(2):134-45. doi: 10.1165/rcmb.2009-0315OC
- 9. Carrier Y, Ma H-L, Ramon HE, Napierata L, Small C, O'toole M, et al. Inter-regulation of th17 cytokines and the il-36 cytokines in vitro and in vivo: Implications in psoriasis pathogenesis.

J Invest Dermatol.2011;131(12):2428-37. doi: 10.1038/jid.2011.234

- Towne JE, Renshaw BR, Douangpanya J, Lipsky BP, Shen M, Gabel CA, et al. Interleukin-36 (il-36) ligands require processing for full agonist (il-36α, il-36β and il-36γ) or antagonist (il-36ra) activity. J Biol Chem. 2011;286(49):42594-602. doi: 10.1074/jbc.M111.267922
- Gresnigt MS, van de Veerdonk FL, editors. Biology of il-36 cytokines and their role in disease. v 2013;25(6):458-65. doi: 10.1016/j.smim.2013.11.003.
- 12. Babaloo AR, Shirmohammadi A, Sandoghchian S, Kamalzadeh A, Ghasemi S. Evaluation of the effect of il- 36γ expression on chronic periodontitis by enhancing the mapk and tlr4 signaling pathways: A basic research. JODDD. 2018;12(3):159. doi: 10.15171/joddd.2018.025
- 13. Carranza FA, Newman MG, Takei H, Klokkevold P. Carranza's clinical periodontology. 10 th ed. Noida: Saunders, Reed Elsevier India Private Limited ; 2006.
- 14. Greenstein G, Hart TC. A critical assessment of interleukin-1 (il-1) genotyping when used in a genetic susceptibility test for severe chronic periodontitis. *J* Periodontol. 2002;73(2):231-47. doi: 10.1902/jop.2002.73.2.231
- Deo V, Bhongade M. Pathogenesis of periodontitis: Role of cytokines in host response. Dent Today. 2010;29(9):60-2, 4-6; quiz 8-9.
- 16. Loos BG. Systemic markers of inflammation in periodontitis. J Periodontol. 2005;76(11):2106-15. doi: 10.1902/jop.2005.76.11-S.2106
- Kurşunlu SF, Öztürk VÖ, Han B, Atmaca H, Emingil G. Gingival crevicular fluid interleukin-36β (-1f8), interleukin-36γ (-1f9) and interleukin-33 (-1f11) levels in different periodontal disease. Arch Oral Biol. 2015;60(1):77-83. doi: 10.1016/j.archoralbio.2014.08.021.
- Garlet G. Destructive and protective roles of cytokines in periodontitis: A re-appraisal from host defense and tissue destruction viewpoints. J Dent Res. 2010;89(12):1349-63. doi: 10.1177/0022034510376402.
- Wang P, Meinhardt B, Andre R, Renshaw BR, Kimber I, Rothwell NJ, et al. The interleukin-1-related cytokine il-1f8 is expressed in glial cells, but fails to induce il-1β signalling responses. Cytokine. 2005;29(6):245-50. doi: 10.1016/j.cyto.2004.12.002
- 20. Schmitz J, Owyang A, Oldham E, Song Y, Murphy E, McClanahan TK, et al. II-33, an interleukin-1-like cytokine that signals via the il-1 receptor-related protein st2 and induces t helper type 2-associated cytokines. Immunity .2005;23(5):479-90. doi: 10.1016/j.immuni.2005.09.015
- 21. Huynh J, Scholz GM, Aw J, Kwa MQ, Achuthan A, Hamilton JA, et al. Irf6 regulates the expression of il-36γ by human oral epithelial cells in response to porphyromonas gingivalis. T J. Immunol. 2016 ;196(5):2230-8.. doi: 10.4049/jimmunol.1501263.
- 22. Sarah S, Tamilselvan S, Kamatchiammal S, Suresh R. Expression of toll-like receptors 2 and 4 in gingivitis and chronic periodontitis. *Indian* J Dent Res .2006;17(3):114.

23. Yamaguchi R, Yoshimura A, Yoshioka H, Kaneko T, Hara Y. Ability of supragingival plaque to induce toll-like receptor 4–mediated stimulation is associated with cytokine production by peripheral blood mononuclear cells. *J* Periodontol.2009;80(3):512-20. doi: 10.1902/jop.2009.080393.

Effect of IL-36y Expression on Chronic Periodontitis 33

24. Kwa MQ, Nguyen T, Huynh J, Ramnath D, De Nardo D, Lam PY, et al. Interferon regulatory factor 6 differentially regulates toll-like receptor 2-dependent chemokine gene expression in epithelial cells. *J* Biol Chem. 2014;289(28):19758-68. doi: 10.1074/jbc.M114.584540.